HIGH PERFORMANCE LIQUID CHROMATOGRAPHY (HPLC) LIQUID CHROMATOGRAPHY / MASS SPECTROMETRY (LC/MS)

Note: Make enough copies of Pages 1-12 to assess each test method in use at the laboratory, one method at a time CHEMISTRY TEST METHOD EVALUATED: Does the laboratory have an in-house methods manual for each accredited analyte or method 5.5.4.1.2(a) Note: This manual may consist of copies of published or referenced test methods Does the laboratory clearly indicate in its methods manual any modifications made to the 5.5.4.1.2(b) referenced test method and describe any changes or clarifications where the referenced test method is ambiguous or provides insufficient detail Does each test method in the in-house methods manual include or reference, where applicable: 5.5.4.1.2(b)(1) **Identification** of the test method 5.5.4.1.2(b)(2) Applicable matrix or matrices **Method Detection Limit** 5.5.4.1.2(b)(3) Scope & application, including components to be analyzed 5.5.4.1.2(b)(4) 5.5.4.1.2(b)(5) Summary of the test method Definitions 5.5.4.1.2(b)(6) 5.5.4.1.2(b)(7) Interferences 5.5.4.1.2(b)(8) Safetv 5.5.4.1.2(b)(9) **Equipment & supplies** 5.5.4.1.2(b)(10) Reagents & standards 5.5.4.1.2(b)(11) Sample collection, preservation, shipment, & storage 5.5.4.1.2(b)(12) **Quality control** 5.5.4.1.2(b)(13) Calibration & standardization 5.5.4.1.2(b)(14) Procedure 5.5.4.1.2(b)(15) Calculations 5.5.4.1.2(b)(16) Method performance 5.5.4.1.2(b)(17) Pollution prevention 5.5.4.1.2(b)(18) Data assessment & acceptance criteria for quality control measures 5.5.4.1.2(b)(19) Corrective actions for out-of-control data 5.5.4.1.2(b)(20) Contingencies for handling out-of-control or unacceptable data 5.5.4.1.2(b)(21) Waste management 5.5.4.1.2(b)(22) References 5.5.4.1.2(b)(23) Tables, diagrams, flowcharts, validation data Does the laboratory ensure that the essential standards outlined in Appendix D are incorporated D into the method manuals and/or Quality Manual

COMMENTS:

 5.5.5.2.2	Do the laboratory's initial & continuing instrument calibration verifications meet the requirements in mandated test methods & regulations (see pages 16-17 for acceptance criteria and the number of standards required)	
	Note: If it is not apparent which standard is more stringent, then the requirements of the regulation or the mandated test method are to be followed	
 5.5.5.2.2.1(a)	Does the laboratory's test method SOP include or reference details of the initial instrument calibration procedures	
	 Note: This includes calculations, integrations, & associated statistics Note: If the test method is referenced for initial instrument calibration procedures, the laboratory must have this method & make it available for review 	
 5.5.5.2.2.1(b)	Does the laboratory retain sufficient raw data records to permit reconstruction of the initial instrument calibration	
	Note: Examples of such data records include calibration date, test method, instrument, analysis date, each analyte name, analyst initials or signature, concentration & response, calibration curve or response factor, and unique equation or coefficient used to reduce instrument responses to concentration	
 5.5.5.2.2.1(c)	Does the laboratory quantitate sample results only from the initial instrument calibration and not from any continuing instrument calibration verifications, unless required by regulation, method, or program	
 5.5.5.2.2.1(d)	 Does the laboratory verify all initial instrument calibrations with a standard obtained from a second manufacturer or lot if the lot can be demonstrated from the manufacturer as prepared independently from other lots Note: When commercially available, traceability shall be to a national standard 	
 5.5.5.2.2.1(e)	Has the laboratory established criteria for the acceptance of an initial instrument calibration Note: Examples include linear regression correlation coefficient & response factor %RSD Note: The acceptance criteria must be appropriate to the calibration technique employed	
 5.5.5.2.2.1(f)	For purposes of establishing the working calibration range , is the lowest calibration standard concentration the lower limit of quantitation	
 5.5.5.2.2.1(f)	Is all data reported below the lower limit of quantitation reported using defined qualifiers or flags or explained in the case narrative	
 5.5.5.2.2.1(g)	Is the highest calibration standard the highest concentration for which quantitative data are to be reported	
 5.5.5.2.2.1(g)	Is all data reported above the highest calibration standard reported using defined qualifiers or flags or explained in the case narrative	
 5.5.5.2.2.1(h)	Does the laboratory report measured concentrations outside the working calibration range as having less certainty & using defined qualifiers or flags or explained in the case narrative	
 5.5.5.2.2.1(h)	Is the lowest calibration standard above the limit of detection for each analyte	

	CHEMI	STRY TEST METHOD EVALUATED:
	Note: F	For instrument technologies (e.g., ICP, ICP/MS) with validated techniques from manufacturers or methods employing standardization with a zero point & a single-point calibration std., the following must occur:
	5.5.5.2.2.1(h)(1)	 Prior to the analysis of samples, are the zero point & single point calibration analyzed, and the linear range of the instrument established by analyzing a series of standards, one of which must be at the lowest quantitation level Note: Sample results within the established linear range will not require data qualifier flags
	5.5.5.2.2.1(h)(2)	Are the zero point & single point calibration standard analyzed with each analytical batch
	5.5.5.2.2.1(h)(3)	Is a standard corresponding to the limit of quantitation analyzed with each analytical batch & meet established acceptance criteria
	5.5.5.2.2.1(h)(4)	Is the linearity verified at a frequency established by the test method and/or the manufacturer
	5.5.5.2.2.1(i)	Does the laboratory perform corrective actions & reanalyze all associated samples if the initial instrument calibration results are outside established acceptance criteria
	5.5.5.2.2.1(i)	 When reanalysis is not possible, does the laboratory report sample data associated with unacceptable initial instrument calibrations with appropriate data qualifiers Note: NELAC Standards 5.5.5.2.2.1(h) & (i) may need to be assessed in conjunction with the Quality Systems data audit
	5.5.5.2.2.1(j)	Does the laboratory have a standard operating procedure for determining the number of points for establishing the initial instrument calibration
	5.5.5.2.2.1(j)	 Does the laboratory use a minimum of two calibration standards (not including blanks or a zero standard) for performing an initial instrument calibration Note: This Standard applies if a reference or mandated method does not specify the number of calibration standards Note: One of the standards must be at the limit of quantitation Note: This Standard does not apply to instrument technologies for which it has been established by methodologies & procedures that a zero & a single point standard are appropriate for calibrations (see Section 5.5.5.2.2.1(h))
COMM	ENTS:	
	5.5.5.10	Does the laboratory verify the validity of the initial calibration by a continuing instrument calibration verification with each analytical batch , prior to sample analyses , whenever an initial instrument calibration is not performed on the day of analysis
	5.5.5.10(a)	Are the details of the continuing instrument calibration verification procedure , calculations , & associated statistics included or referenced in the test method SOP
	5.5.5.10(b)	Is calibration verified for each compound, element, or other discrete chemical species Note: For multi-component analytes such as Aroclors, Total Petroleum Hydrocarbons, or Toxaphene, a representative chemical related substance or mixture can be used

 5.5.5.10(c)(1)	Is the instrument calibration verification performed at the beginning & end of each analytical batch Note: Only one verification needs to be performed at the beginning of the analytical batch if an internal standard is used
 5.5.5.10(c)(2)	Is the instrument calibration verification performed whenever it is expected that the analytical system may be out of calibration or might not meet the verification acceptance criteria
 5.5.5.10(c)(3)	Is the instrument calibration verification performed if the time period for calibration or the most previous calibration verification has expired
 5.5.5.10(c)(4)	Is the instrument calibration verification performed for analytical systems that contain a calibration verification requirement
 5.5.5.10(d)	 Does the laboratory retain sufficient raw data records to permit reconstruction of the continuing instrument calibration verification Note: Such records include test method, instrument, analysis date, name of each analyte, concentration & response, calibration curve or response factor, or unique equations or coefficients used to convert instrument responses into concentrations
 5.5.5.10(d)	Does the laboratory's continuing calibration verification records explicitly connect the continuing verification data to the initial instrument calibration
 5.5.5.10(e)	Has the laboratory established criteria for the acceptance of a continuing instrument calibration verification (e.g. relative percent difference)
 5.5.5.10(e)	Does the laboratory perform corrective actions if the continuing instrument calibration verification results are outside established acceptance criteria
 5.5.5.10(e)	 Does the laboratory perform a new initial instrument calibration if the routine corrective action procedures fail to produce a second consecutive (immediate) calibration verification within acceptance criteria Note: Alternatively, the laboratory can demonstrate acceptable performance after correction with 2 consecutive calibration verifications
 5.5.5.10(e)	 If the laboratory has not verified calibration, do sample analyses not occur until the analytical system is calibrated or calibration verified Note: For sample data associated with an unacceptable calibration verification, the results must be flagged but the data may be useable under the following special conditions: Non-detects for analytes in associated samples where the acceptance criteria for the continuing calibration verifications are exceeded high Any test result for an analyte that indicates exceedence of a maximum regulatory limit or decision level, when the acceptance criteria for the continuing calibration verification for that analyte is exceeded low Any samples with test results that do not meet either of the above criteria must be re-analyzed after a new initial instrument calibration has been established, evaluated, & accepted

COMMENTS:

 5.5.4.2.2(a) C.1	 Has the laboratory performed a satisfactory demonstration of method capability prior to the acceptance & institution of this test method Note: Demonstrations of capability are done in an applicable & available clean quality system matrix sample in a quality system matrix where no target analytes or interferences present at concentrations that impact the results of a specific test method Note: These following steps are may not be applicable for tests with which spiking is not an
	 option and for which Quality Control samples are not readily available Note: Actual sample spike results, such as 4 consecutive matrix spikes (or quality control samples of analytes that do not lend themselves to spiking), within the last 12 months may be used to meet this Standard
	Note: A demonstration of capability is not required in cases where samples are analyzed with this test method in use by the laboratory before July 1999 & where there have been no significant changes in instrument type, personnel, or test method, in which case the analyst's documentation of continued proficiency is acceptable (the laboratory must have records on file to show that a demonstration of capability is not required)
	Note: Continuing demonstration of method performance , per the QC requirements in App. D (e.g., laboratory control samples), is required thereafter
 C.1	Does the laboratory document in its Quality Manual other adequate approaches to Demonstration of Capability if the procedure below is not required by the mandated test method or regulation and if the laboratory elects not to perform this procedure
 C.1 (a)	Is the quality control sample used for this Demonstration of Capability obtained from an outside source
	Note: If an outside source is not available, the laboratory may prepare this sample with stock standards that are prepared independently from those used in instrument calibration
 C.1(b)	Are the analytes diluted in a volume of clean quality system matrix sufficient to prepare 4 aliquots at the specified concentration or to a concentration approximately 1-4 times the limit of quantitation
 C.1(c)	Are at least 4 such aliquots prepared & analyzed according to the test method Note: These analyses may occur either concurrently or over a period of days
 C.1(d)	 Does the laboratory calculate the mean recovery in the appropriate reporting units & the standard deviation of the population sample (n-1) in the same units for each parameter of interest using all of the analysis results obtained Note: When it is not possible to assess mean & standard deviation, such as for presence-absence & logarithmic values, the laboratory must assess performance against established & documented criteria
 C.1(e)	Are the mean and standard deviation for each parameter compared to the corresponding acceptance criteria for precision & accuracy in the test method (if applicable) or in laboratory-generated acceptance criteria (if the method or analyte is non-standard)
 C.1(e)	Does the laboratory consider the performance unacceptable & not analyze actual samples for parameters that fail the acceptance criteria
 C.1(f)	 When one or more parameters fail at least one of the acceptance criteria, does the analyst: Locate & correct the source of the problem, then repeat the test for all parameters of interest, OR Repeat the test for all parameters that failed to meet criteria Note: Repeated failure from employing the second option above indicates a general problem with the entire measurement system, and the analyst must then perform the first option above

 C.1	Is an initial evaluation performed for all analytes to be added to an existing accredited test method (for analytes not currently found on the laboratory's list of accredited analytes)
 5.5.2.6(c)(3)	Does each Analyst have documentation of continued proficiency by at least one of the following once per year:
	 Acceptable performance of a blind sample (single blind to the analyst) An initial measurement system evaluation or another demonstration of capability Successful performance of a blind performance sample on a similar test method using the same technology (acceptable limits must be determined prior to analysis) At least 4 consecutive laboratory control samples with acceptable levels of precision & accuracy (the acceptable limits must be determined prior to analysis) Analysis of authentic samples that have been analyzed by another trained analyst with statistically indistinguishable results
 5.5.4.2.2(d) C.2	Does the laboratory use the NELAC-specified certification statement to document the completion of each Demonstration of Capability (initial & continuing)
 C.2	Are copies of these certification statements retained in the personnel records of each employee performing the test method
 5.5.4.2.2(d) C.1	Does the laboratory retain & make available all associated supporting data necessary to reproduce the analytical results summarized in the appropriate certification statement
 5.5.4.2.2(e) C.1	Does the laboratory complete a demonstration of capability each time there is a change in instrument type, personnel, or test method
 5.5.4.2.2(f)	 Does the laboratory fully document the achievement of demonstration of capability requirements for each specialized work cell Note: A work cell is defined as a group of analysts with specifically defined tasks that together perform the test method
 5.5.4.2.2(g)	Does the laboratory demonstrate & document acceptable performance through acceptable continuing performance checks (e.g, laboratory control samples) each time that membership in a work cell changes
 5.5.4.2.2(g)	Do the new members of the work cell work with experienced analysts in the specialty area
 5.5.4.2.2(g)	Does the laboratory repeat a Demonstration of Capability with the new work cell if the first 4 continuing performance checks following the change in personnel produce a failure in any sample batch acceptance criteria
 5.5.4.2.2(g)	Does the laboratory repeat a Demonstration of Capability if the entire work cell is changed or replaced
 5.5.4.2.2(h)	Is the performance of the work cell as a group linked to the training records of the individual members of the work cell
 5.1.1	 Does the laboratory's procedure for demonstrating its capability to perform the method, the analyst's capability to perform the method, or the acceptance criteria for precision & accuracy comply with the requirements specified in the mandated test method Note: See page 17 for such Demonstration of Capability procedural requirements & acceptance criteria

 D	Does the laboratory have procedures for developing acceptance/rejection criteria for each Chemistry test method (where no regulatory or method criteria exist)
 D	Does the laboratory assess & evaluate all quality control measures on an on-going basis
 D	Does the laboratory use quality control acceptance criteria to determine the validity of the data
 5.5.9.2(d) App. D	Does the laboratory's Chemistry data indicate that the quality control protocols in the test methods manual are being followed (by all analysts)
 5.1.1	Does the laboratory's acceptance criteria for blanks, laboratory control samples, duplicates, & matrix spikes fulfill the requirements in mandated test methods Note: See page 18 for acceptance criteria
 5.1.1	Does the laboratory fulfill additional requirements specified in the mandated test method or regulation Note: See page 19 for the additional requirements stated in test methods
 D.1.1.1(a)	Does the laboratory process the method blank along with & under the same conditions as the associated samples to include all steps in the analytical procedure
 D.1.1.1(a)	Does the laboratory have procedures in place to determine if a method blank is contaminated
 D.1.1.1(b)	Does the laboratory analyze method blanks at a frequency of at least one per preparation batch or one per 20 environmental samples analyzed together with the same method & personnel using the same lots of reagents
 D.1.1.1(c)	Does the method blank consist of a quality system matrix similar to associated samples & known to be free of the analytes of interest
 D.1.1.1(d)	Does the laboratory critically evaluate each method blank as to the nature of any interferences & the effect on the analyses of each sample within the batch
 D.1.1.1(d)	Is the source of the contamination investigated & measures taken to minimize or eliminate the problem
 D.1.1.1(d)	 Are all samples associated with a contaminated blank reprocessed for analysis or reported with appropriate data qualifying codes Note: Such sample results can be reported with data qualifiers: If the analyte concentration in the blank is at or above the reporting limit AND is greater than 1/10 of the amount measured in any sample OR If the method blank contamination affects the sample results as per test method requirements or individual project data quality objectives
 D.1.1.1(d)	Does the laboratory document all corrective actions taken with respect to a contaminated blank

 D.1.1.2(b)	Does the laboratory analyze at least one laboratory control sample (LCS or QC Check Sample) per preparation batch or one per 20 environmental samples analyzed together with the same method & personnel using the same lots of reagents
	Note: This Standard does not apply to analytes for which spiking solutions are not available
	(e.g. Total Suspended Solids, Total Dissolved Solids, Total Volatile Solids, Total Solids,
	pH, Color, Odor, Temperature, Dissolved Oxygen, or Turbidity)
	Note: The matrix spike may be used in place of this control sample as long as the acceptance criteria are as stringent as for the laboratory control sample
	Note: The LCS may consist of media containing known & verified concentrations of analytes or as a Certified Reference Material
 D.1.1.2 (c)	Does the laboratory include all target analytes in the LCS spike mixture over a 2-year period
 D.1.1.2(c)	Are all analyte concentrations in the LCS within the calibration range of the test method
 D.1.1.2(c)	Are the components spiked into the LCS as specified by the mandated test method or other regulatory requirement or as requested by the client
	Note: In the absence of such requirements, the minimum number of analytes to spike are:
	- For methods with 1-10 target analytes, spike all analytes
	- For methods with 11-20 analytes, spike at least 10 analytes or 80%, whichever is greater
	- For methods with more than 20 target analytes, spike at least 16 analytes
	Note: The analytes selected for spiking must be representative of all analytes reported & must
	represent the chemistries and elution patterns of the components to be reported, when some
	components interfere with accurate assessment (e.g., simultaneously spiking technical Chlordane, Toxaphene, & PCB's)
 D.1.1.2 (d)	Does the laboratory document the calculations for percent recovery of the individual batch LCS
 D.1.1.2(d)	Are the individual analyte percent recoveries compared to the acceptance criteria published in the mandated test method or, where such criteria are not established, to client-specified acceptance criteria or to internal criteria determined at the laboratory
	Note: The laboratory must document the method used to establish internal LCS recovery limits
 D.1.1.2(d)	Are all samples associated with an out-of-control LCS reprocessed for analysis or reported with appropriate data qualifying codes
 D.1.1.2 (e)	For large number of analytes in the LCS, does the laboratory take corrective actions if acceptance criteria (3 standard deviations) are not achieved :
	- for 2 analytes when the LCS contains 11-30 analytes
	- for 3 analytes when the LCS contains 31-50 analytes
	- for 4 analytes when the LCS contains 51-70 analytes
	- for 5 analytes when the LCS contains 71-90 analytes
	- for 6 analytes when the LCS contains over 90 analytes
 D.1.1.2(e)	Does the laboratory locate the source of error & take corrective action if the same analyte exceeds LCS control limits repeatedly
 D.1.1.2(e)	Does the laboratory have a written procedure to monitor the application of marginal exceedance allowances to LCS control limits to ensure random behavior

 D.1.1.3	Does the laboratory document procedures for determining the effect of the sample matrix on test method performance
	Note: These procedures relate to the analysis of quality system matrix specific QC samples & could be data quality indicators for a specific sample using a designated test method; these controls alone are not used to judge laboratory performance
 D.1.1.3	Does the laboratory have procedures in place for tracking, managing, & handling matrix- specific QC criteria
	Note: These procedures must include spiking appropriate components at appropriate concentrations, calculating recoveries & relative percent difference, and evaluating & reporting results based on performance of the QC samples
 D.1.1.3.1(b)	Does the laboratory perform matrix spikes (MS) at a frequency specified by the test method Note: This matrix spike analysis frequency is specified in pages xx-xx
	Note: If the test method is not mandated, the laboratory must determine the frequency of matrix spike analysis as part of a systematic planning process (e.g., data quality objectives)
 D.1.1.3.1(c)	Are the components spiked into the MS as specified by the mandated test method or other regulatory requirement or as requested by the client
	Note: In the absence of such requirements, the minimum number of analytes to spike are: - For methods with 1-10 target analytes, spike all analytes
	 For methods with 11-20 analytes, spike at least 10 analytes or 80%, whichever is greater For methods with more than 20 target analytes, spike at least 16 analytes
	 For methods with more than 20 target analytes, spike at least to analytes Note: The analytes selected for spiking should represent the chemistries & elution patterns of components to be reported (e.g., simultaneously spiking Chlordane, Toxaphene, & PCB's)
 D.1.1.3.1(c)	Does the laboratory include all target analytes in the MS spike mixture over a 2-year period
 D.1.1.3.1(d)	Does the laboratory document the calculations for percent recovery & relative percent difference in matrix spikes & matrix spike duplicates
 D.1.1.3.1(d)	Are the individual analyte percent recoveries compared to the acceptance criteria published in the mandated test method
 D.1.1.3.1(d)	If there is no established criteria, has the laboratory determined internal criteria & documented the method used to establish the limits
 D.1.1.3.1(d)	Are all samples associated with matrix spike results outside established criteria documented with corrective actions or reported with appropriate data qualifying codes

COMMENTS:

 D.1.1.3.2(b)	Does the laboratory perform matrix duplicates at a frequency specified by the required mandated test method Note: This matrix duplicate analysis frequency is specified in pages xx-xx
 D.1.1.3.2(c)	Are matrix duplicates performed on replicate aliquots of actual samples
 D.1.1.3.2(d)	Does the laboratory document the calculations for relative percent difference or other statistical treatments
 D.1.1.3.2(d)	Are the individual analyte duplicate precisions compared to the acceptance criteria published in the mandated test method
 D.1.1.3.2(d)	If there is no established criteria, has the laboratory determined internal criteria & documented the method used to establish the limits
 D.1.1.3.2(d)	Are all samples associated with duplicate precisions outside established criteria documented with corrective actions or reported with appropriate data qualifying codes
 D.1.1.3.3(b)	 Does the laboratory add surrogate compounds to all samples, standards, & blanks for all appropriate test methods Note: This Standard does not apply if the sample matrix precludes the use of surrogates or when a surrogate is not commercially available
 D.1.1.3.3(c)	Do the surrogates represent the various chemistries of the method's target analytes & deliberately chosen for being unlikely to occur as an environmental contaminant
 D.1.1.3.3(d)	Are the surrogate recoveries compared to the acceptance criteria in the mandated test method
 D.1.1.3.3(d)	Does the laboratory evaluate surrogate recoveries outside acceptance limits for the effect indicated for the individual sample results
 D.1.5 (a)	Has the laboratory evaluated selectivity by following the checks established within the method Note: These evaluations may include mass spectral tuning, second-column confirmation, chromatography retention time windows, ICP inter-element interference checks, sample blanks, spectrochemical absorption or fluorescence profiles, co-precipitation evaluations, & electrode response factors.
 D.1.5(b)	 Does the laboratory perform confirmations to verify compound identification when positive results are detected on a sample from a location that has not been previously tested by the laboratory Note: These confirmations are performed on pesticides, herbicides, acid extractables, or other organic tests, or when recommended by the analytical test method Note: Confirmation is not required when the analysis involves the use of a mass spectrometer Note: Confirmation is required unless stipulated in writing by the client
 D.1.5(b)	Does the laboratory document all confirmations of compound identity
 D.1.5(c)	If a mass spectrometer is used, has the laboratory documented acceptance criteria for mass spectral tuning

 D.1.2	Does the laboratory document all procedures & retain all supporting data in determining & verifying limits of detection & limits of quantitation
 D.1.2.1	Does this test method provide limits of detection (LOD's) that are appropriate & relevant for the intended use of the data
 D.1.2.1	Has the laboratory determined the limit(s) of detection by the protocol in the mandated test method or applicable regulation
	Note: If the protocol for determining LOD's is not specified, the laboratory must still determine the LOD's but according to a procedure that reflects instrument limitations & intended application of the test method
	Note: In the absence of regulatory or client requirements, an LOD is not required when test results are not reported outside of the calibration range
 D.1.2.1 (a)	Has the laboratory initially determined the detection limits for the compounds of interest in this test method in a quality system matrix in which there are no target analytes or interferences at a concentration that would impact the results
	Note: If this is not possible, the laboratory must determine these detection limits in the quality system matrix of interest
 D.1.2.1(b)	Does the laboratory determine LOD's each time there is a change in the test method that affects how the test is performed or when a change in instrumentation occurs that affects the sensitivity of the analysis
 D.1.2.1(c)	Does the laboratory have established procedures to relate LOD's with Limits of Quantitation (LOQ's)
 D.1.2.1(d)	Has the laboratory verified the LOD annually for each quality system matrix, test method, & analyte
	Note: All sample processing steps of the analytical method must be included in the determination of the LOD
	Note: Validity of the LOD is confirmed by qualitative identification of the analyte(s) in a quality control sample in each quality system matrix containing the analyte at no more than 2-3x the LOD for single-analyte tests and 1-4x the LOD for multiple analyte tests
	Note: LOD verification must be performed on every instrument that is to be used for analysis of samples & reporting of data
	Note: A LOD study is not required for any component for which spiking solutions or quality control samples are not available (e.g., Temperature), or when test results are not to be reported to the LOD (versus the Limit of Quantitation or working range of instrument calibration according to Appendices D.1.2, D.4.5, D.5.4, and D.6.6 to NELAC Chapter 5).

 D.1.2.2(a)	Are all established LOQ's above the LOD's for each analyte	
 D.1.2.2(b)	Has the laboratory verified the LOQ annually for each quality system matrix, test method, & analyte	
	Note: The LOQ study is not required for any component or property for which spiking solutions or quality control samples are not commercially available or otherwise inappropriate (e.g., pH).	
	Note: The validity of the LOQ is confirmed by successful analysis of a quality control sample, containing the analytes of concern in each quality system matrix at 1-2 times the claimed LOQ	
	Note: A successful analysis is one where the recovery of each analyte is within the established test method acceptance criteria or client data quality objectives for accuracy.	
	Note: This single analysis is not required if the bias & precision of the measurement system are evaluated at the LOQ	
	Note: The LOQ verification is not required is not required if the LOD is re-evaluated or verified	
 5.1.1	Do the laboratory's limits of detection fulfill the requirements of mandated test methods or regulations	
	Note: US EPA's Safe Drinking Water Act (SDWA) & Clean Water Act (CWA) regulations require determination of Method Detection Limits according to the procedures & criteria in 40 CFR Part 136, Appendix B	
	Note: See page 18 for SDWA Maximum Contaminant Levels & RCRA Toxicity Characteristics, which the LOD, LOQ, or the lowest-concentration calibration standard must be reliably & consistently below	
	 Note: Other regulations (including state regulations) & permits may contain additional requirements for Reporting Limits, Minimum Levels, Lower Limits of Detection, & other criteria 	

COMMENTS: List analytes for which the above requirements for measurement sensitivity have not been fulfilled

HIGH PERFORMANCE LIQUID CHROMATOGRAPHY (HPLC) LIQUID CHROMATOGRAPHY / MASS SPECTROMETRY (LC/MS)

REQUIRED REAGANTS & STANDARDS

EPA 532 (HPLC-UV) (all HPLC stationary phases are nonpolar reverse-phase, unless otherwise indicated)

Reverse-phase C-18 solid-phase disks or cartridges Ethyl Acetate, Methylene Chloride, Methanol solid-phase conditioning reagents Methanol eluting solvent Sodium Sulfate drying reagent Reverse-phase C18 primary column and Cyano-C18 confirmation column; PO4 buffer mobile phase

EPA 531.1, 531.2 (HPLC-Fluorescence)

o-Phthalaldehyde & 2-Mercaptoethanol post-column derivatizing agents (prepared fresh daily) Sodium Hydroxide post-column hydrolysis reagent Monochloroacetic Acid to adjust sample pH to 2.8-3.2 prior to analysis (EPA 531.1) Potassium Dihydrogen Citrate & Sodium Thiosulfate to adjust sample pH around 3.8 (EPA 531.2) Methanol & water mobile phases, gradient elution (15% to 100% or 10% to 80% Methanol, depending on column (EPA 531.1); Methanol-Acetonitrile-Water 1:0:7 to 1:1:6 to 2:2:4 (EPA 531.2)) Excitation wavelength 340 nm, fluorescence emission wavelength 465 nm

EPA 547, SM6651B (HPLC-Fluorescence)

Cation or anion exchange resin stationary phase o-Phthalaldehyde & 2-Mercaptoethanol post-column derivatizing agents Calcium Hypochlorite post-column oxidizing agent 4% Methanol / 5 mM pH 1.9 Phosphate buffer mobile phase, degassed with helium

EPA 549.2 (HPLC-UV, 308 nm for Diquat)

C-8 solid-phase extraction cartridge or disk
 Methanol, water Cetyl Trimethylammonium Bromide, water, Methanol, water, 1-Hexanesulfonate (Na salt) to condition cartridge
 Methanol, water, Cetyl Trimethylammonium Bromide, water, 1-Hexanesulfonate (Na salt) to condition disk
 Phosphoric Acid / Diethylamine eluting solution
 1-Hexanesulfonic Acid ion-pair concentrate
 Phosphoric Acid / Diethylamine / 1-Hexanesulfonic Acid mobile phase

EPA 550 (HPLC-UV & Fluorescence)

Methylene Chloride extraction solvent Sodium Sulfate drying agent Acetonitrile exchange solvent Acetonitrile & water mobile phases, gradient elution (35% to 100% Acetonitrile)

EPA 550.1 (HPLC-UV & Fluorescence)

Reverse-phase C-18 solid-phase extraction disks or cartridges Methylene Chloride & Methanol to condition disks or cartridges Methylene Chloride eluting solvent (cartridges), or Acetonitrile then Methylene Chloride (disks) Sodium Sulfate drying agent Acetonitrile exchange solvent Acetonitrile & water mobile phases, gradient elution (35% to 100% Acetonitrile)

EPA 555 (HPLC-UV diode array)

Sodium Hydroxide hydrolysis reagent Phosphoric Acid to adjust sample pH<2 after hydrolysis C-18/silica concentrator cartridge + analytical column (solid-phase extraction may also be used) Acetonitrile / Phosphate buffer mobile phase, gradient elution (10% to 90% Acetonitrile)

EPA 605 (HPLC-electrochemical detection)

Chloroform extraction solvent Sulfuric Acid for back-extraction clean-up Methanol exchange solvent 1:1 Acetonitrile/Acetate buffer mobile phase

EPA 629, 631 (HPLC-UV)

EPA 3510, 3520, 3540, 3541 with EPA 8321 & 8325 (LC-MS)

Sulfuric Acid hydrolysis reagent (EPA 631, hydrolyzes Benomyl to Carbendazim at pH<1) Sodium Hydroxide (EPA 631, neutralize sample pH to 6-8, must determine Benomyl by difference) Methylene Chloride extraction solvent Sodium Sulfate drying agent Methanol exchange solvent Surrogate standards (EPA 8000's)

EPA 610, SM6440B (GC-FID or HPLC-UV & Fluorescence); 632, 637, 639 (HPLC-UV)

EPA 3510, 3520, 3540, 3541 with EPA 8100 (GC-FID) or EPA 8310 (HPLC-UV & Fluorescence)

Methylene Chloride extraction solvent Sodium Sulfate drying reagent Acetonitrile exchange solvent (HPLC only) Surrogate standards (EPA 8100, 8310)

EPA 632.1 (HPLC-UV)

Methylene Chloride extraction solvent Sodium Sulfate drying agent 1:1 Acetonitrile/water exchange solvent

EPA 1660 (HPLC-UV)

Acetonitrile extraction solvent Mobile Phase gradient elution (30% Acetonitrile in water to 100% Acetonitrile)

EPA 8315 (HPLC-UV)

Acid to adjust sample pH to 3 Dinitrophenylhydrazine (DNPH) pre-column derivatizing agent Methylene Chloride extraction solvent Sodium Sulfate drying agent Acetonitrile exchange solvent Mobile Phase Gradient elution (60 or 70% Acetonitrile in water to 100% Acetonitrile)

EPA 8318 (HPLC-Fluorescence)

o-Phthalaldehyde & 2-Mercaptoethanol post-column derivatizing agent Methylene Chloride extraction solvent Sodium Sulfate drying agent Methanol exchange solvent Reverse-phase C-18 cleanup cartridge Mobile Phase Gradient Elution (10% CH3OH/CH3CN in H3PO4/H2O to 100% Methanol/Acetonitrile)

EPA 8330 (HPLC-UV)

Acetonitrile extraction solvent Sodium Sulfate drying agent 1:1 Acetonitrile/water exchange solvent Cyano-C18 confirmation HPLC column required along with the reverse-phase column 1:1 Methanol/water mobile phase

EPA 8331, 8332 (HPLC-UV)

EPA 8331 Soil Extraction Solvent & Mobile Phase: 1-Decanesulfonic Acid in Acetic Acid, Methanol, & H2O EPA 8332 Mobile Phase: 60% Acetonitrile in water

EPA 8321 (LC-MS thermospray) & EPA 8325 (LC-MS particle beam)

Ethyl Ether extraction solvent (EPA 8321) Sodium Sulfate drying agent (EPA 8321) Acetonitrile exchange solvent (EPA 8321) MS Tuning Solution (Decafluorotriphenylphosphine Oxide) (EPA 8325) (**daily**) Surrogate standards Mobile Phase Gradient Elution (50% Methanol in water to 100% Methanol, for Organophosphorus Pesticides) (50% Acetonitrile in water to 100% Acetonitrile, for Azo Dyes) (25% to 60% Methanol in Ammonium Acetate Buffer, for Chlorinated Phenoxyacids)

(5% Methanol in Ammonium Acetate Buffer to 100% Methanol, for Carbamates) (25% to 70% Acetonitrile in Ammonium Acetate Buffer, for Benzidines & Organonitrogen Pest.)

EPA 3545 with EPA 8310, 8325

Pressurized Extraction Fluids: 1:1 Methylene Chloride/Acetone, 1:1 Hexane/Acetone, Hexane, or CH2Cl2 **Note:** Exchange solvents are based on clean-up method & determinative methods employed

EPA 3550 with EPA 8310, 8325

Ultrasonic Extraction Fluids: 1:1 Methylene Chloride/Acetone, 1:1 Hexane/Acetone, or Hexane **Note:** Exchange solvents are based on clean-up method & determinative methods employed

EPA 3561 with EPA 8310

Carbon Dioxide supercritical extraction fluid, with Methanol, Water, & Methylene Chloride as modifiers Reconstitution Solvents: 1:1 Acetonitrile/THF or 3:1 Methylene Chloride/Isooctane

EPA 3580 with EPA 8310, 8325

Waste Dilution Solvents: Methylene Chloride or Hexane

EPA 3610, 3611 prior to EPA 8310

Alumina Clean-up Sorbent, conditioned with Hexane 20% Ethyl Ether in Hexane, to elute Phthalate Esters from neutral alumina 30% then 50% Ethyl Ether in Pentane, to elute Nitrosamines from basic alumina 20% Methylene Chloride in Hexane, to elute Di-benzo-p-dioxins & Dibenzofurans from basic alumina 50% Methylene Chloride in Hexane, to elute Di-benzo-p-dioxins & Dibenzofurans from acidic alumina Hexane eluting solvent for Base-Neutral Aliphatics in petroleum waste Methylene Chloride eluting solvent for Base-Neutral Aromatics in petroleum waste Methanol eluting solvent for Base-Neutral Polars in petroleum waste

EPA 610; EPA 3630 prior to EPA 8310

Silica Gel Clean-up Sorbent, activated at 130 C for 16 hours, conditioned with Pentane solvent 40% Methylene Chloride in Pentane, to elute Polynuclear Aromatic Hydrocarbons from silica gel 20% Benzene in Hexane or 100% Hexane, to elute Dibenzo-p-dioxins & Dibenzofurans from silica gel 15% Toluene in Hexane, to elute derivatized Pentachlorophenol from silica gel;

40% then 70% Toluene in Hexane, to elute most Derivatized Phenols from silica gel; then 15% Isopropanol in Toluene, to elute the derivatized Nitrophenols

25% Toluene in Hexane, to elute Derivatized Phenols from silica gel cartridge

Hexane, to elute PCB's, Heptachlor, Aldrin, & DDE from silica gel or silica gel cartridge; then

Methylene Chloride, to elute remaining Organochlorine Pesticides from silica gel; or 50% Ethyl Ether in Hexane, to elute remaining Organochlorine Pesticides from silica gel cartridge

EPA 3640 prior to EPA 8310

Gel Permeation Chrmoatography system with GPC Bio-Beads, UV Detector

GPC Calibration Solution (Corn Oil, Bis(2-ethylhexyl) Phthalate, Methoxychlor, Perylene, Sulfur) (store at 4 C, replace every 6 months)

Methylene Chloride eluting solvent

Semivolatile Organics collected within the Phthalate, Methoxychlor, & Perylene elution times Organochlorine Pesticides & PCB's collected within the Methoxychlor & Perylene elution times

HOLDING TIME, SAMPLE CONTAINER, & SAMPLE PRESERVATION REQUIREMENTS

7 Days to Extract Sample, 7 Days to Analyze Extract; Teflon-lined cap; 4 C; 0.008% Sodium Thiosulfate Benzidines (analyze extract immediately if not stored in oxygen-free system)

7 Days to Extract Sample, 21 Days to Analyze Extract; Amber PVC or Silanized Amber Glass container; 4 C SDWA Diquat

7 Days to Extract Sample, 40 Days to Analyze Extract; glass containers with Teflon-lined cap; 4 C; 0.008% Sodium Thiosulfate Phenols, Haloethers, Benzidines (RCRA), Polynuclear Aromatic Hydrocarbons (CWA)

7 Days to Extract Sample, 40 Days to Analyze Extract; glass containers with Teflon-lined cap; 4 C; 0.008% Sodium Thiosulfate; store in the dark Nitrosamines, Nitroaromatics & Cyclic Ketones, Polynuclear Aromatic Hydrocarbons (SDWA & RCRA)

7 Days to Extract Sample, 40 Days to Analyze Extract; glass containers with Teflon-lined cap; 4 C; pH 5-9

Pesticides (CWA), Organophosphorus Pesticides (RCRA)

14 Days to Analyze Sample, 4 C, Sodium Sulfite & HCl to pH<2 SDWA Chlorinated Acids

14 Days to Extract Sample, 14 Days to Analyze Extract; 4 C SDWA Chlorinated Solvents & Disinfection By-Products

14 Days to Extract or Analyze Sample, 4 C

Other SDWA Pesticides & PCB's

INITIAL INSTRUMENT CALIBRATION ACCEPTANCE CRITERIA FOR MANDATED TEST METHODS

3 standards + blank

SM6020B, 1a & 1b, applies to all SM Organics mtds., int. std. response factor (if used) < 20% RSD
EPA 531.1, 10.2 & 10.3, calibration factor or response factor (if used) < 20% RSD
EPA 547, 9.2
EPA 549.2, 10.3
EPA 550 & 550.1, 9.2 & 9.3, calibration factor or response factor (if used) < 10% RSD
EPA 555, 10.2, calibration factor (if used) < 20% RSD
EPA 605, 610, 7.2 & 7.3, calibration or response factors (if used) < 10% RSD
EPA 629, 631, 632, 637, 639, 7.2 & 7.3, calibration factors or response factors (if used) < 10% RSD
EPA 632.1, 8.2, calibration factor (if used) < 20% RSD
EPA 1660, 7.3, calibration factor (if used) < 20% RSD

5 standards + blank

EPA 8000, 7.4-7.5, calibration factor or response factor (if used) < 20% RSD,

correlation coefficient >0.990 for non-linear calibration

Applies to EPA 8310, 8315, 8316, 8318, 8321, 8325, 8330, 8331, 8332 (HPLC Organics)

Requires **client notification of analytes** quantitated from CF or RF when mtd. criteria not met (and mean RSD < 20%)

EPA 531.2, 532, 10.2, calibration factor (if used) < 30% RSD

CALIBRATION VERIFICATION ACCEPTANCE CRITERIA FOR MANDATED TEST METHODS

Recovery 70-130%

EPA 531.2, 532, 10.3, 50-150% allowed for low-level stds.; CCV also required every 10 samples & end of run

Recovery 75-125% EPA 555, 10.2.3

Recovery 80-120%

EPA 531.1, 10.2.4
EPA 547, 550, & 550.1, 9.4
EPA 549.2, 10.4
EPA 8325, 7.4
SM6020B, 1b (applies to SM Organics methods)
40 CFR 141.40, App. A, 3, mid-range std. plus 60-140% recovery for std. at or below Minimum Reporting Level (applies to SDWA Unregulated Contaminants)

Recovery 85-115% EPA 8000, 7.7

Recovery 90-110% EPA 629, 631, 632, 637, 639, 7.2 & 7.3 EPA 632.1, 8.2 EPA 8331, 7.3.2

PRECISION & ACCURACY ACCEPTANCE CRITERIA FOR MANDATED TEST METHODS (INITIAL DEMONSTRATION OF CAPABILITY)

Method Detection Limit required for each analyte EPA 549.2, 9.3

Mean Accuracy 70-130% for each analyte

EPA 555, 9.3 **EPA 547**, 10.3.2, Precision RSD < 30% from mean

Mean Accuracy 80-120% for each analyte; Precision RSD<20%

EPA 531.1, 9.3, MDL determination also required **EPA 531.2**, **532**, 9.2, MDL study also required & at least 3 days required (not all aliquots extracted same day)

Average Recovery & Standard Deviation of Recovery compared to Acceptance Criteria in Table of Test Method EPA 605, 610, 8.2

EPA 629, 631, 632, 637, 639, 8.2 EPA 1660, 8.2 EPA 8000, 8.6, applicable to all EPA 8000-series methods

QUALITY CONTROL ACCEPTANCE CRITERIA FOR MANDATED TEST METHODS

QC Check Sample Recoveries within 70-130%

EPA 532, 9.6, 50-160% allowed for low-level concentrations **EPA 550 & 550.1**, 10.5.1

QC Check Sample or Matrix Spike Recoveries within the Test Method QC Acceptance Criteria for each Analyte EPA 605, 610, 8.4 EPA 1660, 8.3

External QC Check Sample Analyzed Quarterly

EPA 531.2, 9.11, recoveries 70-130% **EPA 555**, 9.5

Matrix Spike Recoveries within 50-150% or 50-160%

EPA 531.2, 9.8, 70-130% recommended for mid- & high-level spikes, analyzed each analytical batch **EPA 532**, 9.9, 70-130% recommended for mid- & high-level spikes, analyzed each extraction batch

Matrix Spike Recoveries 65-135%

EPA 531.1, 9.7, analyzed every 20 samples or batch

Surrogate Recoveries within 70-130%

EPA 531.2, 532, 9.7

Internal Standards Responses

EPA 531.1, 9.5, 70-130% from last Calibration Verification **EPA 550**, **550.1**, 10.4, 70-130% from last Calib. verification

EPA REGULATORY LEVELS REQUIRING SPECIFIC DETECTION LIMITS

SDWA MAXIMUM CONTAMINANT LEVELS

Benzo(a)pyrene	0.2 ug/L
2,4-D	70 ug/L
Pentachlorophenol	1.0 ug/L
2,4,5-TP (Silvex)	50 ug/L
Dalapon	200 ug/L
Dinoseb	7.0 ug/L
Picloram	500 ug/L
Diquat	20 ug/L
Endothall	100 ug/L
Glyphosate	700 ug/L
Carbofuran	400 ug/L
Oxamyl (Vydate)	200 ug/L

RCRA TOXICITY CHARACTERISTICS

2,4-D	10.0 mg/L
2,4,5-TP (Silvex)	1.0 mg/L

ADDITIONAL REQUIREMENTS

Matrix Spikes analyzed every 10 samples

EPA 549.2, 9.6 EPA 547, 550, 550.1, 10.6 EPA 605, 610, 8.3 EPA 629, 631, 632, 637, 639, 8.4, or Monthly EPA 1660, 8.3 SM6020B, 3c, or Monthly (whichever is more frequent) (applies to all SM Organics methods)

Matrix Spike & Matrix Spike Duplicate each batch of 20 samples or fewer

40 CFR 141.40, App. A, 6, must also **alternate between mid- & low-level** concentrations for spikes (applies to SDWA Unregulated Contaminants)

EPA 8000, 8.5, applies to all 8000-series Organics mtds., may use sample dup. in place of MSD

Quality Control Check Samples analyzed every 10 samples

EPA 605, 610, 8.1.5, frequency may be reduced if Matrix Spike recoveries meet all specified QC criteria **EPA 632.1**, 9.2

Field Duplicates analyzed

EPA 531.2, 9.9, each batch unless MSD analyzed, <50% RSD for low-level, <30% recommended for mid & high level
EPA 532, 9.10, each extraction batch unless MSD analyzed
EPA 632.1, 9.3, ALL samples are collected in duplicate; 10% are analyzed

Peak Gaussian Factor evaluated each analytical batch

EPA 532, 10.2.3, for Flumeturon

GC Retention Time Windows established for each analyte EPA 8000, 7.6

Chromatographic Resolution Checks

EPA 531.2, 9.10 **EPA 8325**, 7.3.2; 3,3'-Dimethylbenzidine & 3,3'-Dimethoxybenzidine

GPC Column Calibration Acceptance Criteria (EPA 3640) (performed weekly)

Retention time shift < 5% compared with the previous calibration Symmetrical peaks observed for all compounds Resolution between Corn Oil, Bis(2-ethylhexyl) Phthalate, Methoxychlor, & Perylene peaks all > 85% Resolution between Perylene & Sulfur peaks > 90% & neither peak is saturated in response