

Use and Interpretation of Quantitative HIV-1 RNA Test Results: Guidance for Laboratories

THIS DOCUMENT OUTLINES THE IMPORTANCE OF ACCURATELY REPORTING RESULTS FROM QUANTITATIVE HIV-1 RNA ASSAYS.

Patient samples that have HIV-1 RNA concentrations that are below the lower limit of quantitation (LLoQ) and above the limit of detection (LoD) of a quantitative HIV-1 RNA assay may be either incorrectly reported by clinical laboratories, incorrectly interpreted by clinicians or both.

Nucleic acid tests (NATs), including qualitative and quantitative HIV-1 RNA assays, can detect HIV-1 RNA approximately 10-12 days after exposure.^{1,2} In comparison, currently available HIV-1/2 antigen/antibody (Ag/Ab) immunoassays have a window period of approximately 15-24 days after exposure, making NATs essential in detecting acute HIV infections.¹

While qualitative HIV-1 RNA assays are used primarily for diagnostic purposes, quantitative HIV-1 RNA assays are typically used to help monitor disease progression and treatment efficacy.

Patient samples that have HIV-1 RNA concentrations below the lower limit of quantitation (LLoQ) and above the limit of detection (LoD) of a quantitative HIV-1 RNA assay may be either incorrectly reported by clinical laboratories, incorrectly interpreted by clinicians or both. Incorrectly reporting and incorrectly interpreting these quantitative HIV-1 RNA results has significant clinical and public health implications.

Virologic Marker of Response to Antiretroviral Treatment

In the United States, antiretroviral therapy (ART) is recommended for all persons with HIV and should be initiated as soon as possible after an HIV diagnosis.³ Quantitative HIV-1 RNA assays, commonly referred to as viral load assays, measure the amount of viral RNA in the blood, which is used as a surrogate marker to monitor the effectiveness of ART and disease progression. An initial quantitative HIV-1 RNA assay is performed prior to ART initiation to establish a “baseline” viral load. While on ART, quantitative HIV-1 RNA assays are performed at regular intervals to measure the viral load. This provides essential information to achieve and maintain viral suppression with the prescribed therapy. Quantitative HIV-1 RNA results must be evaluated in the context of HIV serological results, ART history and other clinical markers to ensure that the determination of HIV status, ART effectiveness and disease progression is accurate. Accurate reporting of these results is essential to preventing transmission of the virus, supporting clinical management and conducting accurate disease surveillance.

There are several assays approved by the US Food and Drug Administration (FDA) for quantitation of HIV-1 RNA in human plasma on automated or semi-automated systems (Table 1) including one with dual intended use claims for both diagnosis and monitoring. Each quantitative HIV-1 RNA assay has specific performance characteristics and specifications including LoD, LLoQ and ULoQ which are included in the instructions for use (Table 1) and depicted visually in Figure 1. While these thresholds are standardized for a particular HIV-1 RNA assay, there may be variability between different assays (usually less than 0.5 log) which laboratories and clinicians should be aware of, particularly for monitoring response to therapy.

Limit of detection (LoD) is the lowest measured concentration of HIV-1 RNA (analyte) that is possible to detect in the test sample with acceptable certainty, typically $\geq 95\%$ under routine laboratory conditions.

Lower limit of quantitation (LLoQ) is the lowest measured concentration of analyte (HIV-1 RNA) that can be quantified within the specified degree of accuracy and precision.

Upper limit of quantitation (ULoQ) is the greatest measured concentration that the analyte/HIV-1 RNA can be quantified within the specified degree of accuracy and precision.

LoD vs LLoQ—What do you need to know?

- The LLoQ can either be equal to or greater than the concentration of the LoD.
When the LLoQ is a greater concentration than the LoD, the challenge of result interpretation and reporting arises. If a sample's concentration falls between the LoD and LLoQ, a quantitative HIV-1 RNA assay is able to detect HIV, but cannot quantify the concentration (Figure 1: Yellow Bar).
- The LLoQ cannot be less than the concentration of the LoD.

Table 1: Reporting Language for Currently Available FDA-Approved Quantitative HIV-1 RNA Tests^{a,b}

Assay & Manufacturer (PMA#)	HIV-1 RNA concentration below LoD	HIV-1 RNA concentration above LoD and below LLoQ	HIV-1 RNA concentration at or above LLoQ and within Linear Range
Result, Interpretation			
Abbott RealTime HIV-1^c Abbott (BP060002)	Not Detected, Target not detected	< 1.60 Log₁₀ or < 40 cp/mL, Detected	1.6-7.0 Log₁₀ 40-10,000,000 cp/mL
Alinity m HIV-1^d Abbott (BP200455)	Not Detected, Target not detected	<LLoQ, Detected <LLoQ	≥ 20 Copies/mL to ≤ULOQ, Detected and quantified (20-10,000,000 cp/mL)
Aptima HIV-1 Quant Dx Assay^e Hologic (BP150318)	Not Detected, HIV-1 RNA not detected	<1.47 Log₁₀ or < 30 cp/mL detected, HIV-1 RNA is detected but at a level below the LLoQ	1.47-7.0 Log₁₀ 30-10,000,000 cp/mL, HIV Concentration is within the linear range of 30 to 10,000,000 cp/mL
cobas HIV-1(6800/8800) Roche (BP150262)	Target Not Detected, HIV-1 not detected	< Titer Min,^f HIV-1 detected, less than (Titer Min)	Titer, (Titer) of HIV-1 Detected (20-10,000,000 cp/mL)
COBAS Ampliprep/COBAS Taqman HIV-1 v.2.0 Roche (BP050069)	Target Not Detected, HIV-1 RNA not detected	< 2.00E+01 cp/mL (20 cp/mL), HIV-1 RNA detected, less than 20 HIV-1 RNA cp/mL	≥ 2.00E+01 cp/mL and ≤1.00E+07 cp/mL (20-10,000,000 cp/mL)

Abbreviations: PMA: Pre-market Approval (per FDA website where the acronym comes from), cp/mL: RNA copies per milliliter

a. Package inserts (also known as “instructions for use” (IFU)) are provided for reference use only. For the most updated version please either login to your account with the manufacturer, review the package insert with the assay that was included with the version that is currently being run, or contact a sales representative.

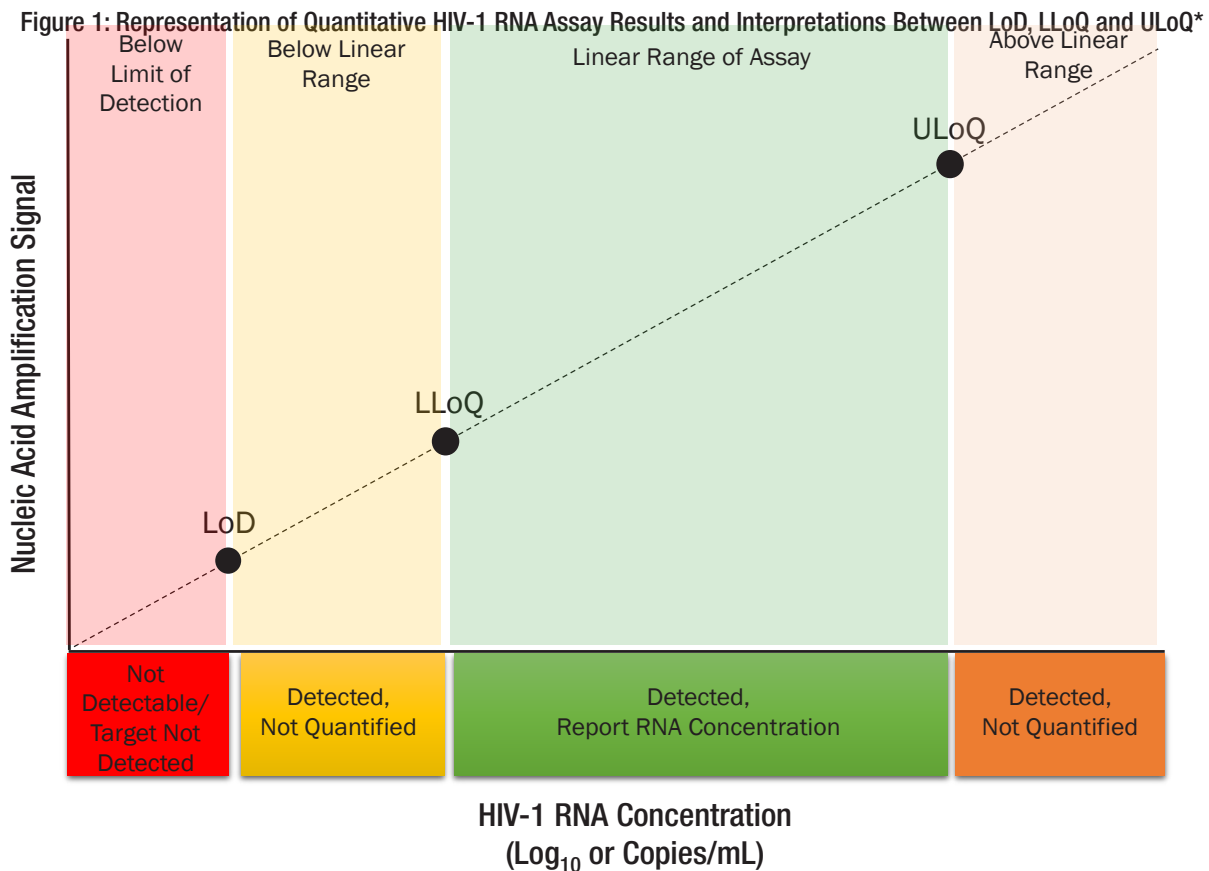
b. Reporting Language provided in the table are based on the IFU for neat or undiluted specimens with regards to the LoD, LLoQ and ULoQ. Please refer to the IFU for the specific assay if using diluted specimens or for further clarification on reporting for each assay.

c. The LoD and LLoQ for the Abbott RealTime HIV-1 Assay varies depending on the sample volume. Reporting Language is for sample volume of 1 mL.

d. The LoD and LLoQ for the Alinity m HIV-1 assay is 20 cp/mL (1.3 log cp/mL) for specimens tested without dilution (neat). The ULoQ is 10,000,000 cp/mL. Diluted specimens (1:25 or 1:50) may be used but if a diluted specimen has an analyte concentration <LoD a message code appears and they should be retested with a new neat specimen, they cannot be interpreted as “target not detected.”

e. The Aptima HIV-1 Quant Dx has dual approved intended uses—one for diagnosis and one for viral load monitoring. This table only includes information for the intended use of monitoring/viral load or quantitation of HIV-1 RNA using plasma, but the assay has an approved intended use as a qualitative assay for the aid in the diagnosis of HIV-1 infection and confirmation of HIV-1 infection. It is intended for use as an aid in the diagnosis of HIV-1 infection, as a confirmation of HIV-1 infection, and as an aid in clinical management of patients infected with HIV-1. See the Instructions for Use for more details.

f. The cobas HIV-1 (6800/8800) uses the term “Titer Min” for LLoQ. Actual Titer Min for the assay is 20 cp/mL for a 500 µL specimen and 50 cp/mL for a 200 µL sample.



* Abbreviations: LoD – Limit of detection; LLoQ – Lower limit of quantitation; ULoQ – Upper limit of quantitation

Reporting Results from Quantitative HIV-1 RNA Assays

Reporting results from quantitative HIV-1 RNA assays is straightforward when HIV-1 RNA is not detected or is detected within (or above) the quantitation range of the assay. When HIV-1 RNA is not detected (either because the RNA concentration is below the LoD or not present), the reporting language may include the terms “not detected,” “target not detected” or “HIV-1 RNA not detected.” When HIV-1 RNA is detected and the concentration is within the linear range of the assay, the specific RNA concentration is reported, alone (Log_{10} cp/mL or cp/mL) or with terms such as: “target detected,” “HIV-1 RNA Detected,” “HIV-1 Detected” (See Table 1, Column 4).

However, reporting and interpreting results from quantitative HIV-1 RNA assays can be confusing if the HIV-1 RNA concentration is greater than the LoD but less than the LLoQ. Samples with an HIV-1 RNA concentration that falls within this zone (Figure 1, yellow bar) are colloquially referred to as “detected, not quantified” or “detectable, not quantifiable” because the assay detects the presence of HIV-1 RNA, but the RNA concentration is lower than the linear range of the assay and, therefore, cannot be quantified reliably. Many manufacturers include a suggested interpretation that is intended to provide more clarity. Specifically for this result, most manufacturers suggest inclusion of interpretations that include the word “detected” (Table 1, Column 3). However, based on reports from US HIV Surveillance Programs, this may not always be reported by the laboratory, may not be consumed properly by the electronic health record (EHR) or may be reported in such a way that the result and result interpretation are not clearly associated with each other.

It is inaccurate and clinically misleading to report or interpret a result of detected, not quantified as HIV-1 RNA negative (target not detected, not detected or undetected). Every effort should be made to ensure that when data are transmitted electronically, details that might clarify the result (i.e., interpretations and comments that accompany the result) are not lost, stripped or otherwise excluded from the version that a healthcare provider reviews to make clinical decisions, as this can lead to misinformed

clinical decisions.

Therefore, APHL recommends that laboratories reporting results from quantitative HIV-1 RNA assays for samples that have an HIV-1 RNA concentration above the LoD but below the LLoQ should make it clear in reporting the result that HIV-1 RNA was detected by including specific language in the result report (i.e., HIV-1 RNA detected, <# copies/mL or HIV-1 RNA Positive/target detected, <#copies/mL) and interpretation (i.e., HIV-1 RNA Detected, Below the Limit of Quantitation for the Assay). Additional language may also be needed to inform providers what additional steps need to be taken.

Table 2: Three examples to illustrate how results and interpretations can vary between different quantitative HIV-1 RNA Assays.^a

	HIV-1 VL Assay A	HIV-1 VL Assay B
LoD	10 copies/mL	10 copies/mL
LLoQ	20 copies/mL	15 copies/mL
Example 1: Sample A has an HIV-1 RNA concentration of 8 copies/mL which is lower than the LoD of both assays and therefore is unlikely to be detected by either assay. It would fall in the range of the red bar in Figure 1.		
Sample A Result	Not Detected	Not Detected
Sample A Interpretation	HIV-1 RNA Not Detected	HIV-1 RNA Not Detected
Example 2: Sample B has an HIV-1 RNA concentration of 15 copies/mL which is greater than the LoD of both assays, lower than the LLoQ for Assay A and equivalent to the LLoQ for Assay B. It should be detected by both assays but will not be able to be quantified by Assay A since it is below the LLoQ. For Assay A it would fall in the range of the yellow bar and with Assay B it would fall in the range of the green bar in Figure 1.		
Sample B Result	<20 copies/mL	15 copies/mL
Sample B Interpretation	HIV-1 RNA Detected at a level below the LLoQ	HIV-1 RNA Detected
Example 3: Sample C has an HIV-1 RNA concentration of 50 copies/mL which is greater than the LoD and the LLoQ for both assays and should be detected and quantified by both assays. This sample would fall within the range of the green bar in Figure 1.		
Sample C Result	50 copies/mL	50 copies/mL
Sample C Interpretation	HIV-1 RNA Detected	HIV-1 RNA Detected

a. Terminology and format for reporting the result and interpretation should follow the manufacturers IFU.

References

1. Delaney KP, Hanson DL, Masciotra S, Ethridge SF, Wesolowski L, Owen SM. Time Until Emergence of HIV Test Reactivity Following Infection With HIV-1: Implications for Interpreting Test Results and Retesting After Exposure. Clin Infect Dis. 2017 Jan 1;64(1):53–9. doi:10.1093/cid/ciw666. Available at <https://academic.oup.com/cid/article/64/1/53/2194435#48974095>
2. Hurt CB, Nelson JAE, Hightow-Weidman LB, Miller WC. Selecting an HIV Test: A Narrative Review for Clinicians and Researchers. Sex Trans Dis. 2017 Dec;44(12): 739-46. doi: 10.1097/OLQ.0000000000000719. Available at https://journals.lww.com/stdjournal/Fulltext/2017/12000/Selecting_an_HIV_Test_A_Narrative_Review_for.5.aspx
3. Guidelines for the Use of Antiretroviral Agents in Adults and Adolescents Living with HIV. Initiation of Antiretroviral Therapy. NIH. Dec 2019. Available at <https://clinicalinfo.hiv.gov/en/guidelines/adult-and-adolescent-arv/initiation-antiretroviral-therapy>

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